

# Journal Pre-proof

Ontogenetic change in the venom composition of one Mexican black-tailed rattlesnake (*Crotalus molossus nigrescens*) from Durango, Mexico

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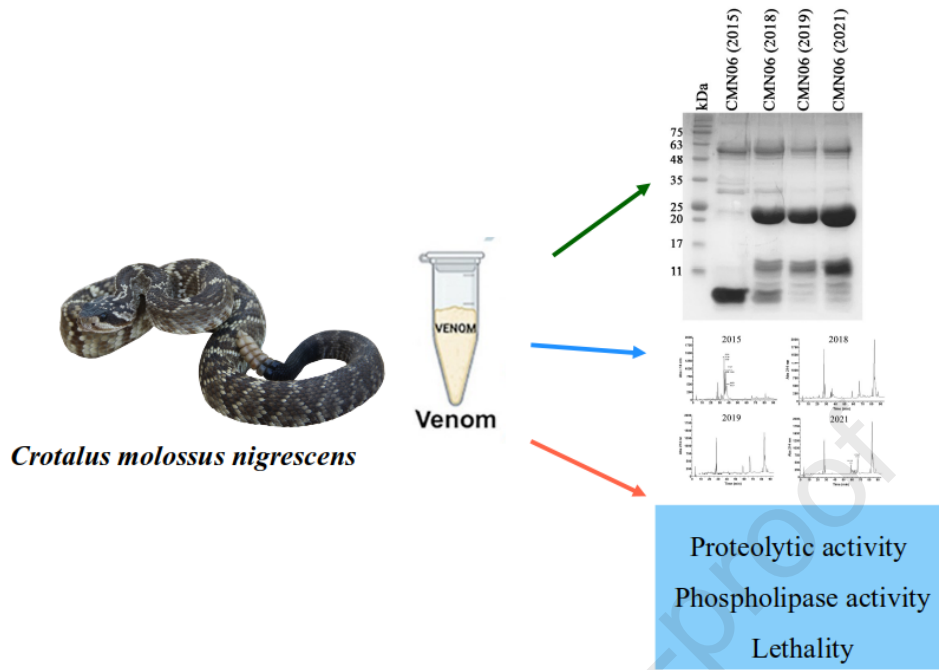
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1 **Ontogenetic change in the venom composition of one Mexican Black-tailed Rattlesnake**  
2 **(*Crotalus molossus nigrescens*) from Durango, Mexico**

3

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20 **Running title:** Ontogenetic venom variation in one *C. m. nigrescens* individual

21

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23 ity; SVMPS; PLA<sub>2</sub>s

24

25 **Abstract**

26 To corroborate the ontogenetic shift in the venom composition of the Mexican Black-tailed  
27 Rattlesnake (*Crotalus molossus nigrescens*) previously reported through the census ap-  
28 proach, we evaluated the shift in the protein profile, lethality, and proteolytic and phospho-  
29 lipase activities of four venom samples obtained in 2015, 2018, 2019, and 2021 from one *C.*  
30 *m. nigrescens* individual (CMN06) collected in Durango, Mexico. We demonstrated that the  
31 venom of *C. m. nigrescens* changed from a myotoxin-rich venom to a phospholipase A<sub>2</sub> and

32 snake venom metalloproteinase-rich venom. Additionally, the proteolytic and phospholipase  
33 activities increased with age, but the lethality decreased approximately three times.

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35 Mexico is the country with the most rattlesnake species (*Crotalus* and *Sistrurus*) in  
36 the world with 47 (Uetz et al., 2023), although few studies about their venom have been  
37 carried out (Neri-Castro et al., 2020). The ontogenetic venom shift has been reported in some  
38 rattlesnake species from Mexico using the census approach (Arnaud et al., 2021; Borja et al.,  
39 2018; Colis-Torres et al., 2022; Mackessy et al., 2018). The “census approach” characterizes  
40 venoms of individuals from different localities and snake sizes (Schonour et al., 2020). How-  
41 ever, these studies have limitations and biases as the venom composition of juveniles and  
42 adults may vary slightly among populations (Margres et al., 2015), reducing the chance to  
43 describe, in detail, the pattern of venom change during one individual’s lifetime.

44 *Crotalus molossus nigrescens* are medium-sized pitvipers distributed broadly in Mex-  
45 ico with a maximum total body length (TBL) of 105 cm (Borja et al., 2018), while newborn  
46 snakes have an average TBL of 27.8 cm (Fernández-Badillo and Torres-Angeles, 2018). Pre-  
47 viously, it has been demonstrated that the venom of *C. m. nigrescens* displays differences in  
48 composition and biological activities in individuals with different TBL (Borja et al., 2018).  
49 For instance, smaller individuals (e.g., TBL<60 cm) contained more crotoamine-like myotox-  
50 ins (MYO) than larger individuals (e.g., TBL>80 cm); however, the opposite occurred with  
51 the snake venom metalloproteinases (SVMPs). In addition, the venoms of the largest indi-  
52 viduals, which contained more SVMPs, were more proteolytic but less lethal than those of  
53 the smallest individuals. Similar results were obtained by Colis-Torres et al. (2022), where  
54 the venom of adult individuals of *C. basiliscus*, a species closely related to *C. m. nigrescens*,  
55 was significantly more proteolytic than the venom of juveniles. To corroborate the ontoge-  
56 netic shift in the venom composition of *C. m. nigrescens* previously reported through the  
57 census approach, we evaluated the shift in the protein profile, lethality, and proteolytic and  
58 phospholipase activity of four samples obtained in different years of the venom from one *C.*  
59 *m. nigrescens* individual collected in Durango, Mexico.

60 To evaluate ontogenetic shifts in the venom composition of *C. m. nigrescens*, we used  
61 venom samples from one female *C. m. nigrescens* individual (CMN06) collected in 2015  
62 from Agua Puerca, Durango, Mexico (26.228341° and -104.492504°) under the collection

63 permit SGPA/DGVS/03562/15 from SEMARNAT. Venom extraction was performed as pre-  
64 viously described by Román-Domínguez et al (2019). Due to samples from years 2016, 2017,  
65 and 2020 being used for other studies and therefore unavailable, we evaluated only four  
66 venom samples from the years 2015, 2018, 2019, and 2021 during which the snake measured  
67 64 cm, 71 cm, 75 cm, and 79 cm snout-vent length (SVL) and a total body length (TBL) of  
68 69 cm, 76 cm, 80 cm, and 84 cm, respectively. Two milligrams of each venom were weighed  
69 and dissolved in 1 mL of phosphate-buffered saline (PBS) pH 7.2 (137 mM NaCl, 2.7 mM  
70 KCl, 10 mM Na<sub>2</sub>HPO<sub>4</sub>, 1.8 mM KH<sub>2</sub>PO<sub>4</sub>). The protein concentration for each sample was  
71 determined using the Bradford assay. The electrophoretic profile of the venom was deter-  
72 mined using 20 µg of venom from each sample dissolved in sample buffer (50 mM Tris-HCl,  
73 pH 6.8, 25% SDS, 10% glycerol, and 0.002% bromophenol blue) and 5% β-mercaptoethanol.  
74 A 12.5% polyacrylamide gel was run under reducing conditions using a discontinuous system  
75 as in Borja et al. (2018). The four venom samples (1 mg in 900 µl of 0.1% TFA) were frac-  
76 tionated by reversed-phase high-performance liquid chromatography (RP-HPLC). RP-HPLC  
77 was carried out on a C-18 analytical column (Agilent, 250 x 4.6 mm) using an Agilent 1100  
78 system as described by Colis-Torres et al. (2022). The protein fractions were detected at 214  
79 nm. We estimated the relative percentage of the most variable fractions of the four samples  
80 using the area under the curve of chromatograms. To determine the molecular mass, we se-  
81 lected the RP-HPLC fractions that displayed notable dissimilarities (in terms of height and  
82 area) among the CMN06 venom samples obtained in 2015 and 2021. ESI-MS obtained the  
83 intact masses on an LCQ Fleet Ion Trap Mass Spectrometer (Borja et al., 2018). The proteo-  
84 lytic activity was assessed using azocasein (Sigma-Aldrich, St. Louis, MO, USA) as sub-  
85 strate, following Colis-Torres et al. (2022). One unit of proteolytic activity was defined as a  
86 change of 0.2 in absorbance per min (Gutiérrez et al., 2008). The synthetic substrate 4-nitro-  
87 3-(octanoyloxy)-benzoic acid (4-NOBA) was used to determine the PLA<sub>2</sub> activity of the  
88 complete venom and of the fractions with a molecular mass expected for PLA<sub>2</sub>. We followed  
89 the procedure described by Holzer and Mackessy (1996) with some modifications. The lethal  
90 potency of the CMN06 venom from 2015 and 2021 was evaluated through LD<sub>50</sub> following  
91 the method initially described by Lorke (1983) with modifications described by Neri-Castro  
92 et al. (2022). Animal experiments were approved by the Bioethics Committee of the Univer-  
93 sidad Nacional Autonoma de Mexico (UNAM) Biotechnology Institute, project 374.

94 Differences in composition were detected in the electrophoretic and chromatographic  
95 profiles of the four venom samples of the CMN06 individual. The most notable changes in  
96 electrophoretic gels were observed in the bands smaller than 17 kDa and the bands between  
97 20 kDa to 25 kDa (**Fig. 1A**). For example, the major band in the venom from the year 2015  
98 was located below 11 kDa (likely containing mostly MYO). However, this band gradually  
99 decreased in intensity in the subsequent years as the snake aged and increased in size. This  
100 result was confirmed by RP-HPLC, where the venom sample from 2015 had four major frac-  
101 tions (representing approximately 69% of the venom (**Table 1**)) that eluted between 30 and  
102 40 min and according to their molecular masses (4959-5187 Da) they correspond to MYO.  
103 These fractions were drastically reduced in height in the sample from 2018 and completely  
104 disappeared in the samples from 2019, and 2021 (**Fig. 1B**). In contrast, the bands with a  
105 molecular mass just above 11 kDa (likely containing phospholipases A<sub>2</sub>s (PLA<sub>2</sub>s) and C-type  
106 lectins (CTLs)) and between 20 kDa and 25 kDa (likely containing PI- and PII-SVMPs and  
107 cysteine-rich secretory proteins (CRISP)) were almost absent in the 2015 venom sample but  
108 increased in intensity in the remaining samples until becoming the dominant bands in the  
109 venom from the year 2021 (**Fig. 1A**). RP-HPLC fractions eluting after 50 min notably in-  
110 creased height and area in the samples from 2018, 2019, and 2021 which corroborated the  
111 SDS-PAGE. Particularly, the intact molecular masses of the fractions that eluted at 58.5 min  
112 (FV), 65.7 min (FVI), and 66.5 min (FVII) were 13,772 Da, 13,813 Da, and 13,797 Da,  
113 respectively (**Fig. 1B**), a molecular mass expected for PLA<sub>2</sub>s. The N-terminus of fraction  
114 FVII (SLVQFEILIMKVAKRSGLFSSAYSAYGCYCGWGGH) confirmed that this fraction is  
115 an acidic PLA<sub>2</sub>. These three fractions comprised approximately 24% of the venom in the  
116 sample from 2021 but only 7.5% of the venom in the sample from 2015 (**Table 1**). The  
117 highest peaks in the venoms from 2018, 2019, and 2021 were eluted after 80 min, with per-  
118 centages of 44.6, 49.6, and 53.6, respectively. In contrast, fractions in this area constituted  
119 only 3.4% of the sample from 2015 (**Table 1**). According to a previous publication (Borja et  
120 al., 2018), these fractions contain mostly SVMPs.

121 The proteolytic activity of the CMN06 venom was  $1.05 \pm 0.03$ ,  $6.94 \pm 0.41$ ,  $7.02 \pm 0.32$ ,  
122 and  $7.05 \pm 0.35$  U/mg for the years 2015, 2018, 2019, and 2021, respectively (**Table 1**). The  
123 phospholipase activity for venoms from 2015, 2018, 2019, and 2021 was  $1.4 \pm 0.4$ ,  $3.1 \pm 0.2$ ,  
124  $3.2 \pm 0.2$ , and  $3.7 \pm 0.2$   $\Delta A$  450 nm, respectively (**Table 1**). The only RP-HPLC fraction with

125 phospholipase activity ( $2.5\pm 0.4$ ) was the one eluting at 66.4 min (FVII). The CMN06 venom  
126 from 2015 ( $LD_{50}$ :  $1.86 \mu\text{g/g}$  (C.I.=1.79-1.93)) was approximately three times more lethal to  
127 mice than CMN06 venom from 2021 ( $LD_{50}$ :  $5.64 \mu\text{g/g}$  (C.I.=5.33-5.96)) (Table 1). In addi-  
128 tion to the shift in lethality, the venom from 2015 generated hind limb spastic paralysis in  
129 mice, an effect that was not observed in mice injected with venom from 2021.

130 In general, two basic venom compositional “strategies” have been observed in rattle-  
131 snakes: venoms with high levels of SVMPs but lower lethality (type I), and venoms with  
132 reduced amounts of SVMPs but higher lethality (type II) (Mackessy, 2010). Our results sug-  
133 gest that *C. m. nigrescens* venom shifts from a type II venom to a type I venom as they grow  
134 up and mature; and support that lethal toxicity and venom metalloproteinase activity are neg-  
135 atively associated (Mackessy, 2010). Crotonamine-like myotoxins (MYOs) are basic peptides  
136 comprised of between 42 and 45 amino acids ( $\sim 4.8$  kDa) (Porta et al., 2021), which induce  
137 myonecrosis and spastic paralysis in the hind limbs of mice, rats, rabbits, and dogs (Gon-  
138 çalves, 1956). Because MYOs rapidly paralyze mammalian prey, it is likely that the high  
139 amounts of this toxin in juvenile *C. m. nigrescens* ensure the rapid immobilization of prey  
140 and prevent escape. Additionally, the high abundance of MYO in juvenile venoms suggests  
141 that this toxin could be responsible for their increased toxicity; however, previously, it has  
142 been shown that the  $LD_{50}$  of MYO is relatively high (1.5-3.0 mg/kg for intravenous (iv))  
143 (Marinovic et al. 2017); therefore, it is likely that several toxins acting in synergy, including  
144 MYO, may induce the higher toxicity in juvenile venoms.

145 PLA<sub>2</sub>s and SVMPs are two of the most abundant components in rattlesnake venoms  
146 (Tasoulis and Isbister, 2017). PLA<sub>2</sub>s have a molecular mass ranging from 13-16 kDa and can  
147 generate diverse biological effects including, neurotoxicity, myotoxicity, cardiotoxicity,  
148 among others (Lomonte and Križaj, 2021). In addition to enzymatic PLA<sub>2</sub>s, viperid venoms  
149 may also contain phospholipase A<sub>2</sub>-like proteins, which lack catalytic activity but induce  
150 myotoxic effects (Lomonte, 2023). SVMPs are classified into three groups (PI, PII, and PIII-  
151 SVMPs) based on the number of domains that they contain. The range of molecular masses  
152 for PI, PII, and PIII-SVMP is 20-30 kDa, 30-60 kDa, and 60-100 kDa, respectively (Olaoba  
153 et al., 2020). Our results suggest that, at least in this individual in particular, the amount of  
154 PLA<sub>2</sub>s, PI and PII SVMPs in venom tends to increase with the age and size, while the ex-  
155 pression of MYO tends to decrease. It has been suggested that, in addition to their



156 contribution to the toxicity of venom, PLA<sub>2</sub>s and PI-SVMPs play a role in digestion (Bernar-  
157 doni et al., 2014; Thomas and Pough, 1979), particularly in environmental temperatures  
158 lower than the optimal for the snake. Although little is known about the diet of *C. m. ni-*  
159 *grescens* (Balderas-Valdivia et al., 2009; Carbajal-Márquez et al., 2023), it is likely that  
160 adults consume large mammal prey, which would be difficult to digest, and the enzymes in  
161 venom may help in the digestive process.

162         Previously, it was reported that the probable threshold size for the shift in the venom  
163 composition of *C. m. nigrescens* is when the snakes reach a TBL close to 70 cm (Borja et al.,  
164 2018). In accordance with the above, the most drastic change in the electrophoretic profile  
165 of the venom of the CMN06 individual was between 69 cm (2015 sample) and 76 cm (2018  
166 sample) of TBL. Nevertheless, the electrophoretic and chromatographic profiles appear to  
167 show a gradual interchange of certain toxin families instead of an abrupt change where some  
168 toxin families completely disappear, and others appear. Unfortunately, we did not evaluate  
169 all the years in this range (e.g., we missed the years 2016, 2017, 2020). This assumption  
170 needs to be tested by evaluating the ontogenetic shift in more individuals from birth to adult-  
171 hood.

172         In conclusion, our results corroborate the differences previously reported in the  
173 venom composition of different *C. m. nigrescens* individuals with different SVL. However,  
174 it is important to note that monitoring the venom composition of more individuals from dif-  
175 ferent geographic regions from birth to adulthood is needed to completely describe the onto-  
176 genetic shift in this species and all the toxin families implied.

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207 **Fig. 1.** SDS-PAGE and RP-HPLC of four venoms from one *C. m. nigrescens* individual

208 (CMN06) milked in the years 2015 (SVL: 64 cm), 2018 (SVL: 71 cm), 2019 (SVL: 75 cm)

209 and 2021 (SVL: 79 cm). **A)** Twenty micrograms of venom were loaded per line in a 15%

210 reducing-SDS-PAGE. A protein ladder was used as a reference to estimate the molecular

211 mass in kDa of venom bands. Note the transition from high to low MYO prevalence and low

212 to high prevalence in SVMPs and PLA<sub>2</sub>s. **B)** One milligram of venom was separated in each

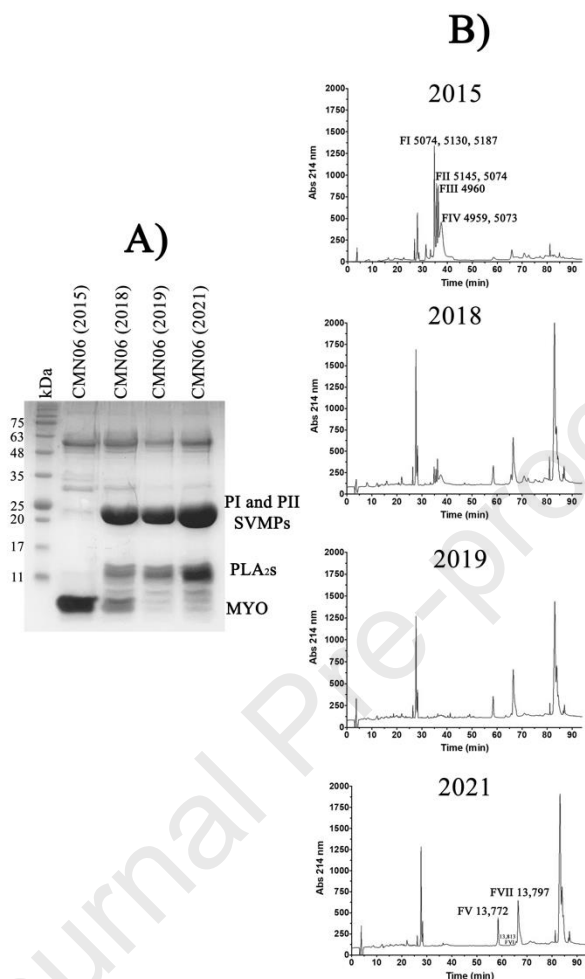
213 run. Proteins were detected at 214 nm and absorbance is indicated on the left axis. The mo-

214 lecular mass (Da) for selected fractions is shown in the chromatograms of venoms from 2015

215 and 2021. Note the reduction of earlier eluting proteins and the increase in later eluting pro-

216 teins over time.

217



218

219 **Table 1.** Relative percentages of PLA<sub>2</sub>s, MYOs, and SVMPs estimated from the area under  
 220 the curve of chromatograms of CMN06 samples from 2015, 2018, 2019, and 2021 and toxic,  
 221 proteolytic, and phospholipase activities for each venom sample.

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Sample	%PLA <sub>2</sub>	%MYO	%SVMPs	Proteolytic activity (U/mg)	Phospholipase activity (ΔA 450 nm)	LD <sub>50</sub> (μg/g)
CMN06 2015	7.4	69.1	3.2	1.05±0.03	1.4 ±0.4	1.86
CMN06 2018	17.6	8.7	44.7	6.94±0.41	3.1±0.2	ND
CMN06 2019	27.6	0	49.6	7.02±0.32	3.2±0.2	ND
CMN06 2021	23.9	1.5	53.6	7.05±0.35	3.7±0.2	5.64

223 ND: Not determined

224

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228

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233

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**Highlights**

The ontogenetic venom shift in *C. m. nigrescens* is reported using a single animal.

Venom composition transitioned from myotoxin-rich to rich in PLA<sub>2</sub>s and SVMPs.

Ontogenetic venom shift predicts proteolytic, phospholipase, and toxic activities.

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## Ethical Statement

The protocols for the use of these animals were authorized by the Institutional Committee for Animal Care at the Biotechnology Institute of the National Autonomous University of Mexico.

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**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

The protocols for the use of these animals were authorized by the Institutional Committee for Animal Care at the Biotechnology Institute of the National Autonomous University of Mexico. We also declare that we have no competing financial interest influencing the work reported on our manuscript.